

## Screening of drugs against Leishmania amazonensis axenic amastigotes in vitro

## Protocol for culturing promastigotes of L. amazonensis (wild type) and differentiation in axenic amastigotes (Vermelho, et al; 2010)

PBHIL Medium (1L): 2g/L glucose 2g/L Peptone 2g/L Brain Heart Broth 0.25g/L Liver Infusion Broth 0.4g/L Sodium chloride 4g/L Potassium chloride 11.5g/L Monobasic sodium phosphate 3g/L Sodium hydroxide 0.01g/L Hemin; 10% FBS.

Add all the ingrediens to 1L of MiliQ water. Separate in 2 beckers of 500ml each. Adjust the pH to 7.2 (promastigotes) and 4.6 (axenic amastigotes). Add 50ml of FBS to each of the beckers and filter.

- 1) Promastigotes culture Grow in PBHIL medium pH 7.2 at 26 °C. Medium can be changed once a week.
- 2) Axenic Amastigotes Spind down promastigotes from stationary phase (5 to 7 days) at 3000rpm for 10 min, break 1. Discard all the supernatant and ressuspend in PBHIL ph 4.6 for counting. Dilute the parasites at 5x10<sup>6</sup> parasites/ml and incubate them at 32-34°C for 5-7 days. Differentiation should be followed by looking in the microscope.

## Protocol for inhibition assay on *L. amazonensis* axenic amastigotes (wild type)

The assay is performed in 96 well transparent sterile plates for 96 hours. Each compound should be tested in duplicate. The controls in triplicate.

Reagents:

PBHIL medium (pH 4.6) Alamar Blue (Sigma)

## Controls:

Medium alone, parasites alone, parasites + 100uM of lonomycin

Protocol:

- Thaw out compounds
- Spin parasites for 10 min at 3000 rpm (9 acceleration, 1 break).
- In the meanwhile, add 100uL of PBHIL medium (pH.4.6) per well in the 96 well plate
- Vortex compounds
- Add 2uL of each compound + 98uL of fresh medium in the first row, pipette up and down to mix and then transfer 100uL to the next row in order to make serial dilutions
- After centrifugation, discard medium by aspiration carefully
- Resuspend parasites in warm medium and count them in Newbauer chamber
- Dilute parasites at 10<sup>7</sup> cells/ml (10<sup>6</sup> cells/well) and plate 100uL per well with multichannel pipette
- Incubate at 32°C for 96h.
- Add 40uL of Alamar blue per well
- Incubate at 32°C for 5 hours
- Read fluorescence at excitation 530nm and 590nm emission wavelength

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